



ANTITUMOUR EFFECTS OF PURE DIASTEREOMERS OF 5-FORMYLTETRAHYDROFOLATE IN HEPATIC TRANSPLANTS OF A RODENT COLON CARCINOMA MODEL

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Abstract—The effects of the two diastereoisomers of 5-formyltetrahydrofolate on tumour growth, thymidylate synthase (TS, EC 2.1.1.45) levels, and potentiation of 5-fluorouracil cytotoxicity were studied in an *in vivo* rat colon carcinoma model, transplanted to liver. The animals were randomized into eight groups, treated with daily i.v. tail vein injections of racemic (d,l)-5-formyltetrahydrofolate (5-CHO-FH₄), 15 mg/kg, (l)-5-CHO-FH₄ 7.5 mg/kg, and (d)-5-CHO-FH₄ 7.5 mg/kg, 5-fluorouracil (FUra) 30 mg/kg, (d,l)-5-CHO-FH₄ 15 mg/kg + FUra 30 mg/kg, (l)-5-CHO-FH₄ 7.5 mg/kg + FUra 30 mg/kg, and (d)-5-CHO-FH₄ 7.5 mg/kg + FUra 30 mg/kg, and a sham-treated control group. The average tumour size of the groups was equal at the start of treatment. After six days' treatment the average tumour sizes were at laparotomy 3.3 ± 1.0 g in the (d,l)-5-CHO-FH₄ treated group, compared to 2.0 ± 0.1 g in the FUra treated group and 7.1 ± 3.1 g in the controls. Natural (l)-5-CHO-FH₄ promoted tumour growth (average tumour weight 10.8 ± 4.0 g), whereas the unnatural (d)-5-CHO-FH₄ alone retarded it (average tumour weight 1.2 ± 0.40 g). (l)-5-CHO-FH₄ induced a significant increase in tumour tissue TS levels by [³H]FdUMP radioligand assay (27.5 ± 8.4 pmol/g tumour tissue) compared to controls (16.8 ± 6.1 pmol/g tumour tissue). Increases in 5,10-methylenetetrahydrofolate and tetrahydrofolate occurred with FUra alone, with a further statistically significant increase in both folates with the addition of (d)-5-CHO-FH₄ to FUra.

Key words: 5-formyltetrahydrofolate; 5-fluorouracil; tumour growth; diastereoisomers; thymidylate synthase

Biomodulation of FUra with folates has in recent years been seen as holding promise for the improvement of the antitumour activity of FUra¹ [1–6]. (d,l)-5-formyltetrahydrofolate (5-CHO-FH₄, folinic acid) is widely used in anticancer therapy as a folate modulator for fluoropyrimidines. The rationale is an increased inhibition of the key *de novo* enzyme thymidylate synthase (TS, EC 2.1.1.45) [7]. The natural substrates for TS are 2'-deoxyuridine 5'-monophosphate (dUMP) and the one-carbon donor 5,10-methylenetetrahydrofolate (CH₂FH₄). The combination of the active nucleotide of FUra, FdUMP, with TS and CH₂FH₄ forms an inhibitory covalent ternary complex that requires CH₂FH₄ in large stoichiometric excess to ensure maximal TS inhibition [8]. A lack of reduced folates is potentially one of the most important factors in explaining the poor efficiency of TS inhibition by FdUMP and resistance to FUra-based chemotherapy [9, 10]. Folate deficiency can be circumvented by the addition of 5-CHO-FH₄ to FUra. 5-formyltetrahydrofolatecalcium (d,l)-5-CHO-FH₄ is an equimolar mixture of the two pteridine ring C₆ diastereoisomers.

It has been stated that only the l-form of 5-formyltetrahydrofolate is biologically active and converted into

active folate cofactors (i.e. 6(l)-CH₂FH₄ [11, 12]). It has recently been reported, however, that the d-isomers are not inert and that the unnatural folates can compete with the l-isomers for intracellular transport and interfere with their intracellular metabolism, acting at various enzymatic levels [13–15].

We have developed a rat colon carcinoma model that has been useful for exploring the *in vivo* pharmacodynamics of FUra/folate therapy [16]. These tumours grow as a well-defined solid tumour mass in the liver, and are repeatedly measurable. Assessment of tumour masses at different stages is used to evaluate the effectiveness of different therapeutic modalities, and the tumours are modestly sensitive to FUra. Growth and pharmacodynamic behaviour of these experimental tumours has proven stable during repeated passages and assays (over 85 passages) [16, 17].

The aim of the present study was to evaluate the effects of the racemic (d,l)-5-CHO-FH₄ and the individual d- and l-forms of 5-CHO-FH₄ in combination with FUra on experimental tumour growth and levels of TS.

MATERIALS AND METHODS

Animals and experimental model

Female Wistar rats were anaesthetised with diethyl-ether inhalation, and the liver exposed via a midline laparotomy. 1.0 × 10⁶ viable cells, trypsinized and immersed in DMEM, of a transplantable N-methyl-N'-nitrosoguanidine-induced tumour (NGW) were inoculated in the subcapsular space of the right liver lobe. Inoculation of precise numbers of cells was done by trypsinization followed by negrosin staining and viability counting in a Bürker chamber of freshly passaged cells. Tumour

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|| Abbreviations: NGW, nitrosoguanidine induced adenocarcinoma; FH₄, H₄-pteroylglutamate; CH₂FH₄, 5,10-CH₂H₄-pteroylglutamate; 5-CHO-FH₄, 5-formyl-H₄-pteroylglutamate (i.e. leucovorin); FUra, 5-fluorouracil; TS, thymidylate synthase; and SHMT, serine hydroxymethyltransferase.

take was 100%. The NGW tumour is a rapidly growing poorly differentiated colon carcinoma with a volume doubling time of approximately 24 hours. Relaparotomy was performed at day eight and the longest (A) and the perpendicular (smallest) (B) diameters of the liver tumours were calculated according to the approximation formula,

$$V = A \times B^2 \times 0.5.$$

Treatment was given by tail vein as bolus injections over 2 minutes, daily, days 9 to 11 and 14 to 16. Treatment was always given between 9:00 and 11:00 a.m. to avoid effects of diurnal variation [18]. 5-CHO-FH₄ preparations were given in the same vein as a bolus injection within 30 seconds after FUra.

The animals were killed at day 17. The body weight and tumour volume were registered, the tumour mass excised, and the tumour wet weight registered. Tumour biopsies were stored at -70°C until the analyses of TS and reduced folates were performed.

Experimental design

One hundred twenty rats were inoculated with the NGW tumour in the liver, and then randomized at day 8 to the eight different treatment groups. One group (*n* = 15 animals) received (d,l)-5-CHO-FH₄ 15 mg/kg; one group (*n* = 25 animals) received FUra 30 mg/kg; one group (*n* = 15 animals) FUra 30 mg/kg, and (d,l)-5-CHO-FH₄ 15 mg/kg; one group (*n* = 10 animals) received (l)-5-CHO-FH₄ 7.5 mg/kg; one group (*n* = 10 animals) received (d)-5-CHO-FH₄ 7.5 mg/kg; one group (*n* = 10 animals) received FUra 30 mg/kg, and (l)-5-CHO-FH₄ 7.5 mg/kg; one group (*n* = 10 animals) received FUra 30 mg/kg, and (d)-5-CHO-FH₄ 7.5 mg/kg; and the control group (*n* = 25 animals) received sham injections of saline 0.1 mL. There were no significant differences in the day 8 tumour volume or body weight among the eight experimental groups when the treatment started (Table 1).

Chemicals

FUra, (d,l)-5-CHO-FH₄, (l)-5-CHO-FH₄, and (d)-5-CHO-FH₄ were supplied by Lederle Cyanamid, Wayne, NJ. Pure diastereoisomers of 5-CHO-FH₄ were 99% free of the other diastereoisomer. The doses of FUra and 5-CHO-FH₄ were chosen according to initially performed animal toxicity testing to achieve maximal antitumour effect without lethality to the rats.

L. Casei TS as an ammonium sulfate preparation (Biopure, Boston, MA) was stored at -20°C with an

equal amount of glycerol. Serial dilution of enzyme for the TS and folate assays was done in homogenization buffer (see below), supplemented with 0.2% BSA. The initial concentration was 1200 mIU/mL.

[6-³H]FdUMP (specific activity of 18–20 Ci/mmol) was purchased from Moravsek Biochemicals®, Brea, CA. Purification of radiolysis breakdown products was carried out monthly by DEAE-column separation [16].

Homogenization buffer (100 mL) was prepared by adding 550 mg 5'-CMP, 420 mg sodium fluoride, 28 µL 2-mercaptoethanol, 1 g Na-ascorbate (pH 7.0), and 200 µg BSA to 0.18 M Tris-HCl buffer, pH 7.4. *Folate buffer* was prepared by adding 52.8 µL of 14.3 M 2-mercaptoethanol, 26.4 µL of 37% (w/w) formaldehyde, and 22.752 mL of homogenization buffer, pH 7.4, to 25 mg (d,l)-FH₄ (Sigma, St. Louis, MO). The mixture was incubated for 10 min at room temperature (in the dark), which allowed completion of CH₂FH₄ formation. All chemicals used for preparation of these buffers were purchased at the highest available grade from different commercial sources.

Analysis of thymidylate synthase and reduced folates

The TS binding activity and concentration of CH₂FH₄ and FH₄ in the NGW tumour was measured at day 17, 24 hours after the last drug injection.

Portions of frozen tumour tissue were weighed and minced with scissors in a 10-fold excess of homogenization buffer at 4°C. Homogenates were prepared by the action of rotating shearing blades (Turrax) for 30 sec at 4°C.

The [³H]FdUMP ligand-binding assay was used for analysis of free TS [19]. The objective was to compare intratumoural free TS levels in the drug administered to the control animals, and subsequently analysis of total TS in the tumour specimens was not performed. One mL of the homogenate (duplicate samples) was centrifuged at 4°C and 4000 × *g* for 20 min. To 50 µL of the cytosol was added 25 µL of homogenization buffer, 50 µL [³H]FdUMP (approximately 200,000 dpm) in H₂O, and 25 µL of 46.2 µmol CH₂FH₄. These procedures were carried out at 2–4°C. The tubes were incubated at 37°C for 10 min, then 3 mL of 3% acid charcoal (rapidly stirred at 4°C) was added. This was followed by vortex mixing and centrifugation at 4°C and 4000 × *g* for 15 minutes to precipitate unbound radioactivity; then 0.8 mL of the supernatant was subjected to beta scintillation counting. TS concentrations are expressed as pmole bound FdUMP per gram tissue (wet weight).

Tumour tissue levels of CH₂FH₄ and FH₄ were assayed by adopting the methodology of Priest with some modifications [8, 19–21]. To 50 µL aliquots of cytosol or homogenization buffer alone (blank) were added 50 µL of the [³H]FdUMP (approximately 20 pmole or 800,000 dpm), 4 pmole of *L. Casei* TS (×1.7 = 6.8 pmole of FdUMP binding sites) in 25 µL of homogenization buffer (with 0.2% BSA), and 25 µL of folate buffer (above), with or without formaldehyde for estimation of FH₄ + CH₂FH₄ and 5,10-CH₂H₄, respectively. These procedures were carried out at 2–4°C. The tubes were then incubated at 37°C for 10 min, and 1 mL of ice-cold 3% acid charcoal was added to separate protein-bound radioactivity. The samples were centrifuged at 4°C and 4000 × *g* for 15 minutes, and scintillation counting performed. Folate concentrations are expressed as nanomoles per gram tissue (wet weight).

Table 1. Animal weights and tumour volumes at start of drug therapy at day 8 (means ± SD; *n* = number of rats)

Treatment group	<i>n</i>	Animal weight (g)	Tumour volume (mm ³)
Control	25	199 ± 2.6	261 ± 77.0
FUra	25	198 ± 2.5	260 ± 151.0
d,l-5-CHO-FH ₄	15	200 ± 2.3	261 ± 77.2
FUra + d,l-5-CHO-FH ₄	15	199 ± 1.7	259 ± 86.2
l-5-CHO-FH ₄	10	200 ± 4.4	261 ± 47.9
FUra + l-5-CHO-FH ₄	10	198 ± 4.2	250 ± 72.4
d-5-CHO-FH ₄	10	198 ± 3.2	257 ± 57.5
FUra + d-5-CHO-FH ₄	10	198 ± 3.4	261 ± 30.5

Statistics

Standard parametric test (i.e. Student's *t*-test), one-way ANOVA with the Scheffe *F*-test, and regression analysis were performed to describe and compare data using the StatView software package. Data are presented as arithmetic means \pm standard deviations (SD). *P* \leq 0.05 was considered statistically significant.

RESULTS

Antitumour effects

Tumour wet weights are presented in Fig. 1. Tumour volumes (Table 2) at day 17 correlated with tumour weight ($r = 0.896$, Fig. 1). The tumours in rats treated with FUra were significantly smaller than in the controls, 2.03 ± 0.98 g compared to 7.09 ± 3.08 g. When FUra and (d,l)-5-CHO-FH₄ were given together, the inhibition of tumour growth was more pronounced, but not statistically significant compared to FUra alone. The tumours in the animals treated with (d,l)-5-CHO-FH₄ only were significantly smaller than the control tumours (3.29 ± 0.959 g), although the growth inhibition induced by 5-formyltetrahydrofolate alone was not as pronounced as the FUra effect.

l-5-CHO-FH₄ had a statistically significant growth-stimulating effect (10.81 ± 4.03 g). In contrast, single agent d-5-CHO-FH₄ had a statistically significant growth-inhibiting effect *per se*, with a mean tumour weight of 0.60 ± 0.20 g. Addition of l-5-CHO-FH₄ to FUra increased growth inhibition, but this did not reach statistical significance compared to FUra alone, nor did the addition of d-5-CHO-FH₄ to FUra.

Drug toxicity

No mortality was associated with any of the treatments. Rats treated with FUra therapy showed epiphoria (tearing or excess of lacrimation), whereas the untreated control animals showed no sign of health deterioration, despite increasing tumour masses. Adverse clinical signs could not be observed in the animals treated with (d,l)-5-CHO-FH₄ or (l)-5-CHO-FH₄ only, despite the lack of weight gain in the (l)-5-CHO-FH₄-treated group (Tables

Table 2. Response and toxicity of the treatment, animal weight, and tumour volume at termination of drug therapy at day 17 (means \pm SD). *n* = number of surviving rats at the end of the experiment. Therapy was given days 9–11 and 14–16 by means of rapid bolus i.v. tail injections. All comparisons were made to control rats, and * indicates statistical significance (*P* \leq 0.05).

Treatment group	<i>n</i>	Animal weight (g)	Tumour volume (mm ³)
Control	25	209 \pm 4.1	6307 \pm 1571
FUra	25	179 \pm 11.3*	2184 \pm 1591*
d,l-5-CHO-FH ₄	15	208 \pm 3.0	5916 \pm 2485
FUra + d,l-5-CHO-FH ₄	15	187 \pm 2.5*	1678 \pm 1100*
l-5-CHO-FH ₄	10	198 \pm 9.6	7691 \pm 4266
FUra + l-5-CHO-FH ₄	10	173 \pm 11.5*	1291 \pm 783*
d-5-CHO-FH ₄	10	187 \pm 2.9*	1027 \pm 428*
FUra + d-5-CHO-FH ₄	10	170 \pm 8.9*	475 \pm 157*

1 and 2). (d)-5-CHO-FH₄ was mildly toxic as judged by the animal weight at day 17, which was significantly lower than the control animal weight.

TS levels

In the control animals, TS levels averaged 16.8 ± 6.14 pmole/g. In the animals treated with (l)-5-CHO-FH₄, the TS levels were significantly increased (27.5 ± 8.36 pmole/g; see Table 3). For the other treatments, the differences in TS levels compared to controls were not statistically significant, although the free TS levels were decreased after treatment with FUra. The combination of FUra and (d)-5-CHO-FH₄ increased intracellular CH₂FH₄ concentrations significantly compared to the controls, 5.64 ± 5.50 vs 0.31 ± 0.56 nmole/g, and increased FH₄ and FH₄ levels as well.

DISCUSSION

In the present study we have found that (d,l)-5-CHO-FH₄ retards tumour growth in rats (Fig. 1). The pharmacokinetics of (l)-5-CHO-FH₄ and (d)-5-CHO-FH₄ differ completely, with $\beta_{1/2}$ of 7.5 hours and 32 minutes,

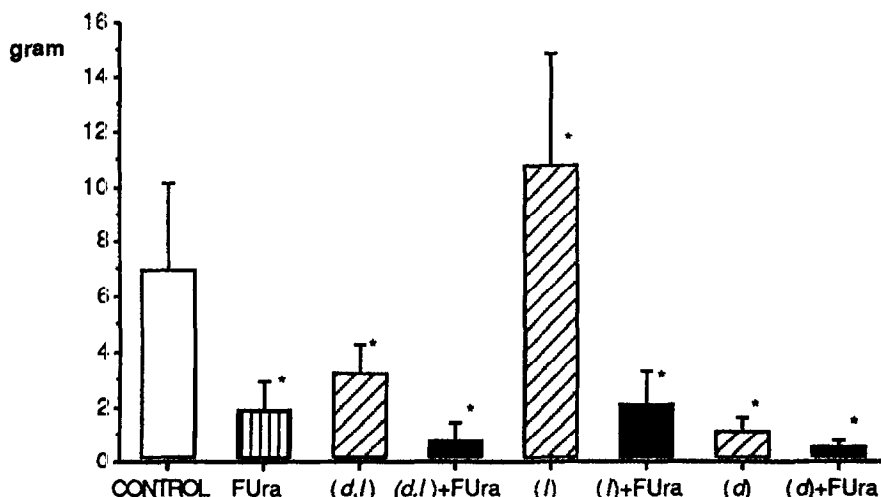


Fig. 1. Tumour weight at day seventeen in the eight groups after termination of drug treatment (means \pm SD). * indicates statistically significant difference (*P* $<$ 0.05) compared to the control group. (d,l) = (d,l)-5-CHO-FH₄ (15 mg/kg), (l) = (l)-5-CHO-FH₄ (7.5 mg/kg), (d) = (d)-5-CHO-FH₄ (7.5 mg/kg). White bars—control, striped bar—5-fluorouracil (FUra, 30 mg/kg), hatched bars—folate only, black bars—folate in combination with FUra (30 mg/kg).

Table 3. Tissue levels (means \pm SD) of TS, CH₂FH₄, FH₄, and total concentrations of reduced folates in tumour tissue at day 17, 24 hours after last drug injection. All comparisons were made to control rats, and * indicates statistical significance ($P \leq 0.05$).

Treatment	TS pmole/g	Folates nmole/g		
		CH ₂ FH ₄	FH ₄	[CH ₂ FH ₄ + FH ₄]
Control	16.8 \pm 6.14	0.38 \pm 0.60	1.34 \pm 1.33	1.72 \pm 1.88
FUra	12.5 \pm 4.66	2.06 \pm 2.82	2.27 \pm 1.25	4.33 \pm 3.85
(d,l)-5-CHO-FH ₄	13.2 \pm 2.35	0.84 \pm 1.11	1.72 \pm 0.85	2.57 \pm 2.06
FUra + (d,l)-5-CHO-FH ₄	10.5 \pm 4.04	0.75 \pm 0.85	1.57 \pm 1.48	2.31 \pm 2.38
(l)-5-CHO-FH ₄	27.5 \pm 8.36*	0.48 \pm 0.79	2.26 \pm 1.05	2.74 \pm 1.42
FUra + (l)-5-CHO-FH ₄	10.1 \pm 5.95	1.07 \pm 0.89	2.01 \pm 0.94	2.91 \pm 1.77
(d)-5-CHO-FH ₄	18.0 \pm 5.07	1.07 \pm 1.23	1.87 \pm 0.60	2.94 \pm 1.72
FUra + (d)-5-CHO-FH ₄	12.2 \pm 5.29	5.64 \pm 5.50*	1.88 \pm 0.85	7.52 \pm 5.58*

respectively, in human plasma for the two isomers [22]. Significant differences in pharmacokinetics of the two isomers have also been reported in animals [23]. Thus, repeated exposure will result in increasing plasma levels of the unnatural isomer. (d)-5-CHO-FH₄ is less efficiently transported into cells than (l)-5-CHO-FH₄, but it has been shown that (d)-5-CHO-FH₄ acts as a competitive inhibitor of (l)-5-CHO-FH₄ transport [24].

The mechanisms of the direct antitumour effect of (d,l)-5-CHO-FH₄ fall outside the scope of this study. Although (d,l)-5-CHO-FH₄ has been widely used clinically for more than 30 years, its metabolic role is not yet completely known. Recent studies by Stover and Schirch [25] indicate that naturally occurring (l)-5-CHO-FH₄ is the result of conversion of 5,10-methenyltetrahydrofolate by serine hydroxymethyltransferase (SHMT, EC 2.1.2.1). In turn, (l)-5-CHO-FH₄ is a slow tight binding inhibitor of the latter enzyme. The active folate interacting with TS and FdUMP is CH₂FH₄, and the currently accepted theory for 5-CHO-FH₄ enhancement of FUra treatment efficacy is expansion of intracellular CH₂FH₄ and/or FH₄ pools [26, 15]. In mice, CH₂FH₄ and pools are elevated 250 to 400%, which supports this hypothesis, but elevation of the reduced folate pool does not account for the inhibition of tumour growth in animals not treated with FUra. However, the expanded folate pools decline rapidly, and approach control levels six hours after 5-formyltetrahydrofolate injection [23].

(l)-5-CHO-FH₄ comprises 30% of the intracellular folate pool, and is an important inhibitor of many folate dependent enzymes. A complex enzyme system exists intracellularly for interconversion of folates and the formation of CH₂FH₄ from exogenous (d,l)-5-CHO-FH₄, but it is still not known if ((d)-5-CHO-FH₄ and (l)-5-5-CHO-FH₄ are metabolised to CH₂FH₄ with the same efficiency. Polyglutamates of (d,l)-5-CHO-FH₄ have been reported to inhibit several enzymes involved in one-carbon donor metabolism, and these inhibitory effects are restricted to the polyglutamates. 5,10-methenyltetrahydrofolate synthase (EC 6.3.3.2), which has been isolated from human liver [27], is the only known enzyme that uses (l)-5-CHO-FH₄ as a substrate. 5,10-methenyltetrahydrofolate synthase is strictly stereoselective across species, and there is no basis to suppose that (d)-5-CHO-FH₄ could be metabolised intracellularly. Using MCF-7 human breast cancer cells, Bertrand and Jolivet found that inhibition of methenyltetrahydrofolate synthetase by 5-formyltetrahydrohomofolate resulted in a two-fold increase in cellular (d,l)-5-CHO-FH₄ concentration with a concomitant and equal decrease in cellular 5,10-methenyltetrahydrofolate concentration. It was also

found that the increased cellular levels of (d,l)-5-CHO-FH₄ were correlated with decreases in both cell growth and rate of *de novo* purine biosynthesis [28].

It was further reported that (l)-5-CHO-FH₄ inhibits *E. coli* TS noncompetitively versus CH₂FH₄ [29, 30]. Although (d)-5-CHO-FH₄ is a considerably less potent TS inhibitor compared to (l)-5-CHO-FH₄ + FUra *in vitro*, it may influence natural folate interactions with TS, since the inhibitory effect of (d,l)-5-CHO-FH₄ is five times lower compared to (l)-5-CHO-FH₄ [13].

(l)-5-CHO-FH₄ has shown itself to be an effective tumour growth stimulator in this study. This growth stimulatory effect was also associated with a significant increase in intratumour TS activity. TS levels were measured 24 hours after the last 5-formyltetrahydrofolate injection, and only (l)-5-CHO-FH₄ was associated with a significant increase in all eight treatment groups. One reasonable explanation for this could be an induction of TS gene expression [10, 13], but decreased TS catabolism potentially could occur. Increases in TS and (l)-CH₂FH₄ after (l)-5-CHO-FH₄ might be expected, *a priori*, to be associated with increased growth rate.

In this study the most pronounced tumour growth inhibition was surprisingly found after administration of (d)-5-CHO-FH₄ and (d)-5-CHO-FH₄ in combination with FUra. Only limited data are available regarding the effects of (d)-folates on folate dependent enzymes, and since the intracellular metabolism of (d)-5-CHO-FH₄ is unknown, the biochemical rationale for this remains to be explained. The antitumour effect of (d,l)-5-CHO-FH₄ may be largely due to the unnatural (d)-5-CHO-FH₄, and it may be hypothesized that the enhancement of FUra cytotoxicity is not a potentiation but an additive effect of (d)-5-CHO-FH₄, since (l)-5-CHO-FH₄ did not significantly enhance the FUra-effect nor expand the folate pools in this study. In a previous report by O'Connell, it was suggested that low-dose (d,l)-5-CHO-FH₄ (20 mg/m²) results in higher response rates than a higher dose (200 mg/m²) in combination with FUra [31]. One explanation for this could be that the cytotoxic effects of (d)-5-CHO-FH₄ predominate at lower concentrations, whereas the tumour growth stimulatory effects of (l)-5-CHO-FH₄ dominate at higher concentrations. We have recently found, when studying the effects of the different diastereoisomers of CH₂FH₄ in a rat colon adenocarcinoma, that unnatural (d)-CH₂FH₄ seems to be an inhibitor of tumour growth *per se*, whereas modulation of FUra activity is the main effect of (l)-CH₂FH₄, which by itself acts as a tumour promoter [32].

In this study we have reported results *in vivo* showing that the biological effects of the two different diastereo-

isomers of 5-CHO-FH₄ seem to have completely opposite effects as single agents. Treatment with unnatural (d)-5-CHO-FH₄ alone was associated with slower tumour growth, whereas natural (l)-5-CHO-FH₄ promoted tumour growth.

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